

FIVE MIKANOLIDE DERIVATIVES FROM *MIKANIA CYNANCHIFOLIA* AND THEIR BIOGENETIC RELATIONSHIPS

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Key Word Index—*Mikania cynanchifolia*; Compositae; sesquiterpene lactones; lactones related to mikanolide; biogenetic relationships.

Abstract—The investigation of the aerial parts of *Mikania cynanchifolia* afforded, in addition to miscandenin, five new sesquiterpene lactones, miscandenin-1,2-epoxide, an isomiscandenin and three further dilactones related to mikanolide. The structures of these new compounds were elucidated by high field ^1H NMR spectroscopy. The biogenetic relationships of the sesquiterpene lactones are briefly discussed.

INTRODUCTION

So far chemical investigations of representatives of the large genus *Mikania* (tribe Eupatorieae, subtribe Mikaniinae) have shown that highly oxygenated sesquiterpene lactones like mikanolide [1–4] and miscandenin [1] may be characteristic for this genus though also several other sesquiterpene lactones [5–10] and a few different types of diterpenes have been reported from several species. We now have studied the constituents of *Mikania cynanchifolia* Hook. et Arn.. The results are discussed in this paper.

RESULTS AND DISCUSSION

The aerial parts of *M. cynanchifolia* afforded a complex mixture of sesquiterpene lactones which could only be separated by repeated TLC and HPLC. In addition to miscandenin (6) [1] five new lactones were obtained, the isabelin derivatives 1 and 2, the isomer of miscandenin 7, miscandenin-1,2-epoxide (8) and the 11 β ,13-dihydro derivative of 3 β -acetoxyisabelin (9). The structures of 1 and 2 clearly followed from the ^1H NMR spectral data (Table 1) which were close to those of one of the congeners of isabelin [11]. The presence of an additional oxygen function was indicated by low field signals at δ 5.08 and 5.95 respectively. Spin decoupling showed that these function were at C-3, while the couplings observed indicated their β -orientation. The ^1H NMR spectrum of 9 (Table 1) was close to that of 2. The presence of an 11 β , 13-dihydro derivative followed from the additional methyl doublet at δ 1.44 which replaced the methylene signals in the spectrum of 2. Of course the H-7 signal was also shifted upfield and the additional double quartet at 2.57 was assigned to H-11. The large coupling $J_{7,11}$ clearly showed that the 11-methyl group was α orientated. The ^1H NMR spectra of all three isabelin derivatives showed that due to the additional oxygen function at C-3 these lactones are present in only one conformation.

The molecular formula of 7 was $\text{C}_{15}\text{H}_{14}\text{O}_5$ and therefore this lactone was formal an isomer of mis-

Table 1. ^1H NMR spectral data of compounds 1, 2 and 9 (400 MHz, CDCl_3 , TMS as internal standard)

	1	2	9
H-1	5.24 <i>br dd</i>	5.13 <i>br dd</i>	5.07 <i>br d</i>
H-2	2.80 <i>ddd</i>	2.87 <i>ddd</i>	2.93 <i>ddd</i>
H-2'	2.35 <i>dd</i>	2.45 <i>br d</i>	2.42 <i>br d</i>
H-3	5.08 <i>ddd</i>	5.95 <i>br dd</i>	5.95 <i>br dd</i>
H-5	7.17 <i>t</i>	7.00 <i>t</i>	7.00 <i>t</i>
H-6	5.31 <i>dd</i>	5.31 <i>dd</i>	5.11 <i>br d</i>
H-7	3.28 <i>dddd</i>	3.30 <i>dddd</i>	2.45 <i>dd</i>
H-8	4.49 <i>ddd</i>	4.47 <i>ddd</i>	4.39 <i>ddd</i>
H-9	3.10 <i>br dd</i>	3.13 <i>br dd</i>	3.09 <i>br dd</i>
H-9'	1.93 <i>dd</i>	1.96 <i>dd</i>	1.89 <i>dd</i>
H-11	—	—	2.57 <i>dq</i>
H-13	6.48 <i>d</i>	6.50 <i>d</i>	1.44 <i>d</i>
H-13'	5.88 <i>d</i>	5.87 <i>d</i>	
H-14	1.61 <i>br s</i>	1.62 <i>br s</i>	1.62 <i>s br</i>
OAc	—	2.15 <i>s</i>	2.17 <i>s</i>

J (Hz): Compounds 1 and 2: 1, 2 = 12.5; 1, 2' = 2', 3 = 2; 2, 2' = 14; 2, 3 = 4; 3, 5 = 5, 6 = 1.5; 6, 7 = 1.5; 7, 8 = 9; 7, 13 = 3; 7, 13' = 3; 8, 9 = 4.5; 8, 9' = 11; 9, 9' = 14; compound 9: 1, 2 = 12.5; 1, 2' = 2', 3 = 2; 2, 2' = 14; 2, 3 = 4.5; 3, 5 = 5, 6 = 6, 7 ~ 1.5; 7, 8 = 9, 5; 7, 11 = 13; 8, 9 = 4; 8, 9' = 11; 9, 9' = 14; 11, 13 = 7.

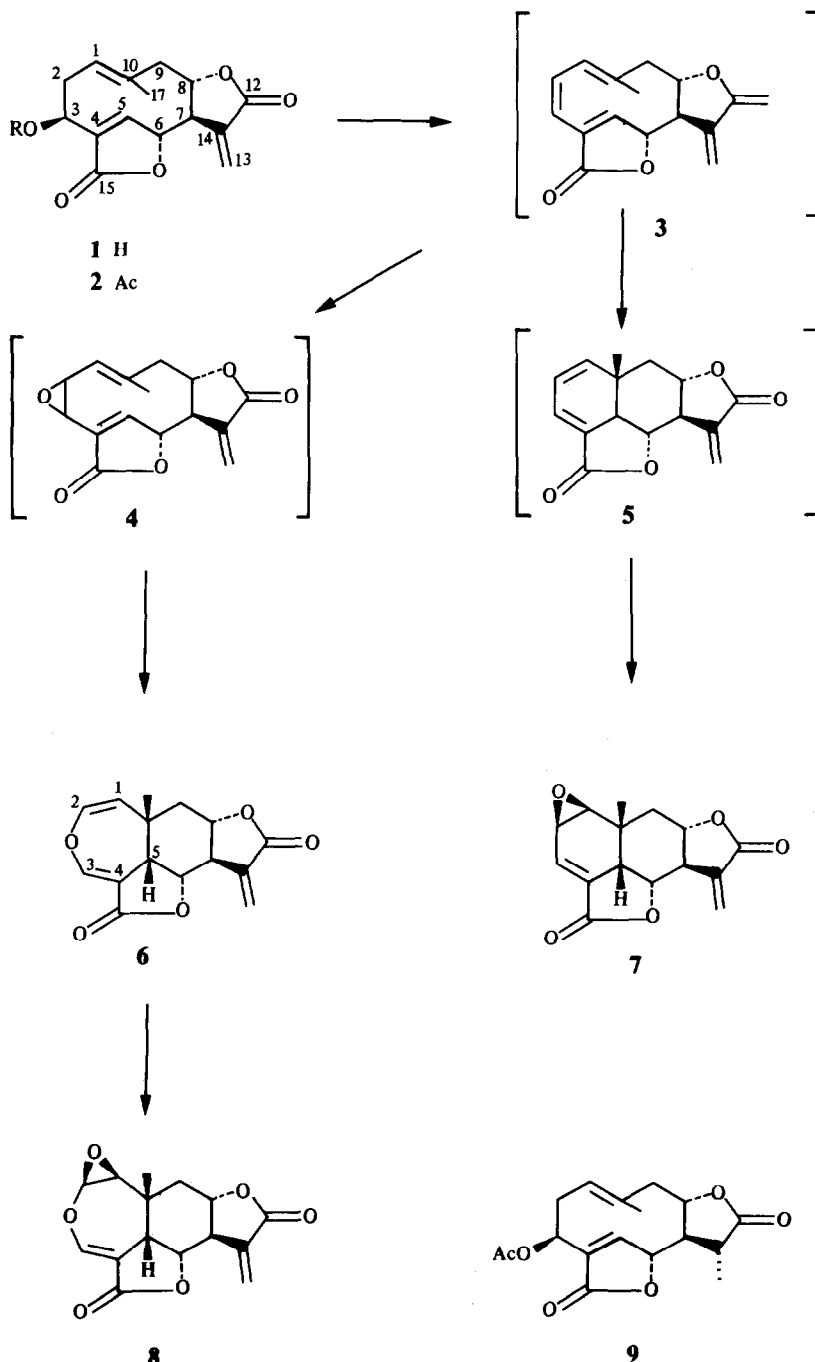
candenin (6). Though several ^1H NMR signals also were similar to those of 6 (Table 2), signals at δ 3.31 and 3.69 required the presence of an epoxide. The chemical shift of an olefinic proton was at δ 7.19 and, contrary to the corresponding signal in the spectrum of 6, this proton had two coupling partners. Spin decoupling showed that it was coupled with the epoxide proton (3.69 *dd*) and with the proton which gave rise to a double doublet at 2.91. As the latter was further coupled with one of the protons under a lactone oxygen, the structure of 7 finally was established. The stereochemistry at C-1 and C-2 was

deduced from the chemical shift of H-14 which could be explained only by a deshielding effect of a β -orientated oxygen function. We have named 7 mikacynancholide.

The structure of 8, molecular formula $C_{15}H_{14}O_6$, again followed from the characteristic 1H NMR spectrum (Table 2) which was in part close to that of 6. However, again the presence of an epoxide was indicated by a doublet at $\delta 2.93$ ($J = 3$ Hz) which was coupled with a doublet at 5.15. As the low field doublet of H-3 still was present (7.23 δ) the epoxide must be at C-1/C-2. Again the

downfield shift of H-14 indicated a β -epoxide. This could be established by epoxidation of 6 which only gave 8 and inspection of a model showed that the *cis*-anellation of the rings strongly favoured a β -attack of the peracid. As the structure of miscandenin was established by X-ray analysis [12] the structure and the configuration of the new epoxide was confirmed.

As shown in Scheme 1 the biogenetic relationships of the lactones present in this *Mikania* species are obvious. Compound 1 is surely formed by allylic oxidation of



Scheme 1.

Table 2. ^1H NMR spectral data of compounds 6–8 (400 MHz, CDCl_3 , TMS as internal standard)

	6	7	8
H-1	4.80 <i>d</i>	3.31 <i>d</i>	2.93 <i>d</i>
H-2	6.17 <i>d</i>	3.69 <i>dd</i>	5.15 <i>d</i>
H-3	7.53 <i>d</i>	7.19 <i>dd</i>	7.23 <i>d</i>
H-5	3.51 <i>dd</i>	2.91 <i>dd</i>	3.30 <i>dd</i>
H-6	4.76 <i>dd</i>	4.93 <i>dd</i>	4.80 <i>dd</i>
H-7	2.72 <i>dddd</i>	2.35 <i>dddd</i>	2.66 <i>dddd</i>
H-8	4.00 <i>ddd</i>	4.07 <i>ddd</i>	4.09 <i>ddd</i>
H-9	2.22 <i>dd</i>	2.16 <i>dd</i>	2.26 <i>dd</i>
H-9'	1.92 <i>dd</i>	1.62 <i>dd</i>	1.98 <i>dd</i>
H-13	6.23 <i>d</i>	6.26 <i>d</i>	6.26 <i>d</i>
H-13'	5.90 <i>d</i>	5.93 <i>d</i>	5.91 <i>d</i>
H-14	1.31 <i>s</i>	1.54 <i>s</i>	1.51 <i>s</i>

J (Hz): Compound 7: 1, 2 = 2, 3 = 3, 5 = 3.5; 5, 6 = 7; 6, 7 = 10; 7, 8 = 11; 7, 13 = 7, 13' = 3; 8, 9 = 3; 8, 9' = 11; 9, 9' = 12; compound 8: 1, 2 = 3, 5 = 3; 5, 6 = 6.5; 6, 7 = 10; 7, 8 = 11; 7, 13 = 7, 13' = 8, 9 = 3; 8, 9' = 9, 9' = 11.5.

isabelin and would lead to 3 by elimination of water. Epoxidation of the 2,3-double bond would give 4, while Cope-rearrangement of 3 could lead to 5. Epoxidation of the latter would give mikacynancholide (7) and a Cope-rearrangement [1] of 4 would give miscandenin (6) as discussed earlier [13].

EXPERIMENTAL

The air dried aerial parts (130 g) (collected in the province Bahia, Brazil, voucher RMK 8913, deposited in the US National Herbarium, Washington) were extracted with Et_2O –petrol, 1:2 (12 hr, room temp.), and the extract obtained was worked-up in the usual fashion. CC fractions (50 ml) were as follows: 1 (petrol), 2 (Et_2O –petrol, 1:10), 3 (Et_2O –petrol, 1:3 and 1:1), 4 (Et_2O and Et_2O –MeOH, 10:1). TLC of fraction 4 (SiO_2 , PF 254, Et_2O –petrol, 3:1, detection of zones by UV light, 255 nm) afforded mixtures of 1, 2 and 6–9 (R_f 0.5) which by TLC (Et_2O – C_6H_6 – CH_2Cl_2 , 1:1:1) and HPLC (RP 8) (MeOH– H_2O , 3:1) finally gave 5 mg 1 (R_f 4.5 min), 20 mg 2 (R_f 7.5 min), 2 mg 7 (R_f 6.9 min), 3 mg 8 (R_f 6.7 min) and 10 mg 9 (R_f 7.3 min). Quantities were determined by weighing.

3 β -Hydroxyisabelin (1). Colourless crystals (not sufficient to obtain a sharp mp but homogeneous by TLC in several solvent systems); IR $\nu_{\text{max}}^{\text{CCl}_4}$ cm^{-1} : 3600 (OH), 1780 (γ -lactone); MS m/z (rel. int.): 276.100 $[\text{M}]^+$ (5) ($\text{C}_{15}\text{H}_{16}\text{O}_5$), 258 $[\text{M} - \text{H}_2\text{O}]^+$ (2), 112 (100), 97 $[\text{112} - \text{Me}]^+$ (58).

$$[\alpha]_{24}^{25} = \frac{589}{+4} \frac{578}{+4} \frac{546}{+2} \frac{436}{-11} \text{CHCl}_3; c = 0.15$$

3 β -Acetoxyisabelin (2). Colourless crystals, mp 163.5°; IR $\nu_{\text{max}}^{\text{CCl}_4}$ cm^{-1} : 1775 (γ -lactone), 1750 (OAc); MS m/z (rel. int.): 318.110 $[\text{M}]^+$ (47) ($\text{C}_{17}\text{H}_{18}\text{O}_6$), 276 $[\text{M} - \text{ketene}]^+$ (100), 258 $[\text{M} - \text{HOAc}]^+$ (14), 206 (44), 164 (56), 119 (52), 111 (56).

$$[\alpha]_{24}^{25} = \frac{589}{+7} \frac{578}{+8} \frac{546}{+7} \frac{436}{-9} \text{CHCl}_3; c = 0.3$$

Mikacynancholide (7). Colourless gum, which could not be induced to crystallize, though no impurities were detectable by TLC in several solvent systems. IR $\nu_{\text{max}}^{\text{CCl}_4}$ cm^{-1} : 1780 (γ -lactone); MS m/z (rel. int.): 274.084 $[\text{M}]^+$ (16) ($\text{C}_{15}\text{H}_{14}\text{O}_5$), 256 $[\text{M} - \text{H}_2\text{O}]^+$ (6), 245 $[\text{M} - \text{CHO}]^+$ (10), 217 $[\text{245} - \text{CO}]^+$ (9), 69 (100).

1 β ,2 β -Epoxy miscandenin (8). Colourless crystals, mp 228.5°; IR $\nu_{\text{max}}^{\text{CHCl}_3}$ cm^{-1} : 1770 (γ -lactone); MS m/z (rel. int.): 290.079 $[\text{M}]^+$ (47) ($\text{C}_{15}\text{H}_{14}\text{O}_6$), 261 $[\text{M} - \text{CHO}]^+$ (100), 246 $[\text{M} - \text{CO}_2]^+$ (70), 216 (20), 109 (63), 69 (71).

$$[\alpha]_{24}^{25} = \frac{589}{-167} \frac{578}{-172} \frac{546}{-198} \frac{436}{-353} \text{CHCl}_3; c = 0.2$$

10 mg 6 in 2 ml CH_2Cl_2 were stirred in the presence of a drop of satd NaHCO_3 solution for 10 hr with 10 mg *m*-chloroperbenzoic acid. TLC (C_6H_6 – Et_2O – CH_2Cl_2 , 1:1:1) afforded 7 mg 6 and 2 mg 8, identical with the natural products (^1H NMR, TLC).

3 β -Acetoxy-11 β ,13-dihydroisabelin (9). Colourless crystals, mp 236–237°; IR $\nu_{\text{max}}^{\text{CCl}_4}$ cm^{-1} : 1780 (γ -lactone), 1750 (OAc); MS m/z (rel. int.): 320.126 $[\text{M}]^+$ (40) ($\text{C}_{17}\text{H}_{20}\text{O}_6$), 278 $[\text{M} - \text{ketene}]^+$ (52), 260 $[\text{M} - \text{HOAc}]^+$ (32), 208 (77), 166 (40), 152 (100), 93 (80).

$$[\alpha]_{24}^{25} = \frac{589}{0} \frac{578}{6} \frac{546}{-10} \frac{436}{-14} \text{CHCl}_3; c = 0.1$$

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